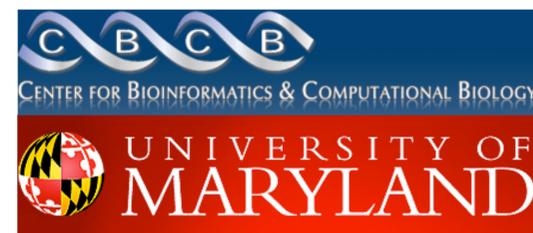


Better Modules in Protein-Protein Interaction Networks

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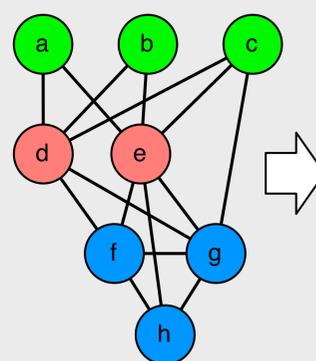
Module-assisted Prediction

A technique called Graph Summarization (GS, ref. 2) can be used to partition protein-protein interaction networks to reveal modules that are more biologically relevant than the clusters produced by other graph partitioning techniques. We apply GS to predict Gene Ontology annotations of biological process for proteins of unknown annotations. We also apply it to detecting membership in protein complexes, as annotated in the MIPS catalog. GS outperforms other approaches such as MCODE, MCL and modularity.

Graph Summarization Example

GS produces a compressed **summary graph**, with nodes combined into supernodes and superedges representing bicliques, and a **list of corrections** to the summary.

Goal: minimize # edges in the summary plus # of corrections.

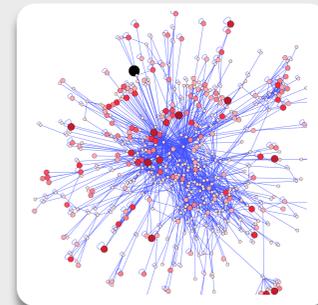


Original PPI network

Graph Summarization Summary

Add {c,g}
Subtract {d,h}

Corrections to the Summary



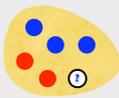
GS of yeast PPI network.
Before: 5,492 proteins
After: 1632 supernodes.

Transferring Annotations

We decompose the PPI network into modules using several graph clustering approaches: MCODE [4], MCL [3], modularity [5], and Graph Summarization [1,2].

We transfer annotations within these modules using one of three standard transfer techniques. Performance is assessed with leave-one-out cross validation.

Majority: transfer if > 50% annotated proteins have the annotation.



Plurality: transfer the most common annotation(s).



Hypergeometric: transfer the statistically enriched annotations.

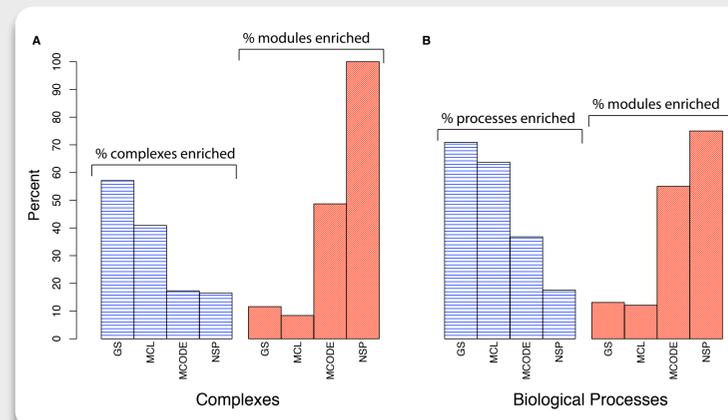


Results: Module Quality

GS modules cover more

At right: the % of annotations statistically enriched in at least 1 module (blue) and the % of modules that are enriched for at least one annotation (red). A wider variety of annotations are enriched in some GS module.

Modularity creates only 8 modules and MCODE clusters only 6% of the network. A larger % of their modules are enriched for some annotation, but this is a poor indicator of predictive performance.



Results: Predicting New Annotations

Interaction network

The PPI network for *Saccharomyces cerevisiae* was downloaded from IntAct (5,492 proteins, 40,332 edges). Similar results are obtained if only edges with ≥ 2 supporting publications are included.

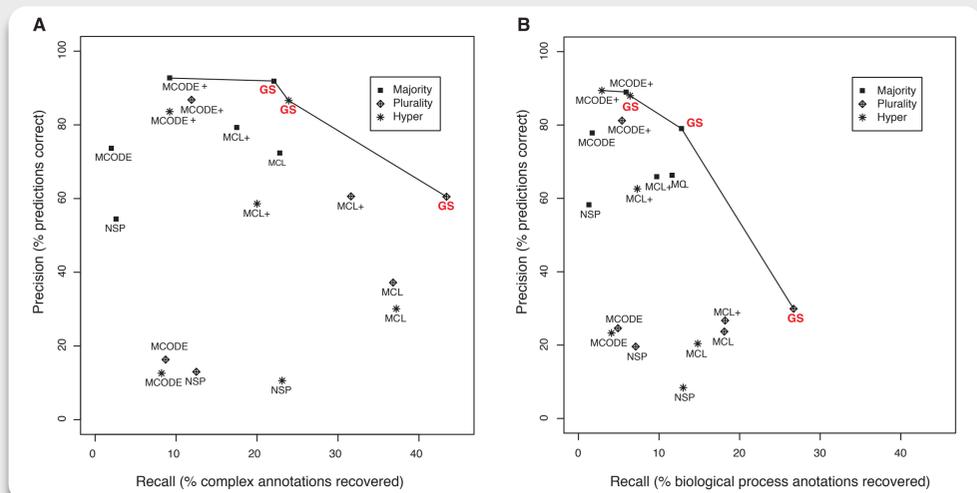
Known annotations

GO biological process annotations for the terms selected in [7] were taken from the SGD. Protein complex annotations were taken from the lowest level of the MIPS complex hierarchy.

GS outperforms MCODE, modularity, MCL

Precision and recall on leave-one-out cross validation is shown using 3 different module-detection methods and 3 different annotation transfer schemes. MCL and MCODE have several parameters. Results are reported for both their default values and parameters tuned to maximize precision (marked with a +). GS has no parameters to tune.

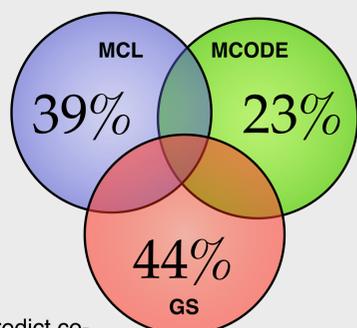
Graph Summarization is always Pareto optimal: no other method dominates it in both precision and recall. Though MCODE can be tuned to achieve precision equal to GS, it has lower recall.



Results: Comparing Predictions

Methods complement each other

A large % of correct predictions are unique to each method using the majority transfer rule and MIPS complex annotations. Similar results hold for biological process annotations and other transfer rules.



Predicting co-complexed pairs

The GS correction list can also be used to predict co-complexed pairs of proteins, with generally higher precision than the method of completing defective cliques [6].

Conclusions

GS produces better biological modules

GS groups together proteins with similar interaction partners, leading to better complex and biological process modules than MCODE, MCL or modularity.

MDL principle

GS is mathematically well-founded and has no parameters to tune.

GS is versatile

It generalizes bipartite cores, cliques, stars. It also explicitly handles noise via a corrections list.

References

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