



Celera Assembler

Theory and Practice

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Celera Assembler Overview



- Primarily developed in 25 man years by 13 computer scientists at Celera for the private human genome effort.
- Attacks repeats by screening high copy repeats, finding repeat boundaries, and utilizing mate-pair information.
- Currently available as an open source project:
<http://wgs-assembler.sourceforge.net>



CENTER FOR
BIOINFORMATICS &
COMPUTATIONAL BIOLOGY

Celera Sequencing Factory



The DNA is loaded into automated sequencers. Celera's automated sequencers run 24-7 and have the ability to decipher more than 100 million letters of genetic code per day - the equivalent of 3 percent of the entire human genetic code every day.

The sequencers create an image of the DNA samples being decoded. The four letters of the genetic code -- A, C, T, G -- each are assigned a color.





Celera Sequencing Factory



- **300 ABI 3700 DNA Sequencers**
- **50 Production Staff**
- **20,000 sq. ft. of wet lab**
- **20,000 sq. ft. of sequencing space**
- **800 tons of A/C (160,000 cfm)**
- **\$1 million / year for electrical service**
- **\$10 million / month for reagents**



Human Data (April 2000)



- Collected 27.27 Million reads = 5.11X coverage
- 21.04 Million are paired (77%) = 10.52 Million pairs
 - 2Kbp 5.045M 98.6% true * <6% std.dev.
 - 10Kbp 4.401M 98.6% true * <8% std.dev.
 - 50Kbp 1.071M 90.0% true * <15% std.dev.

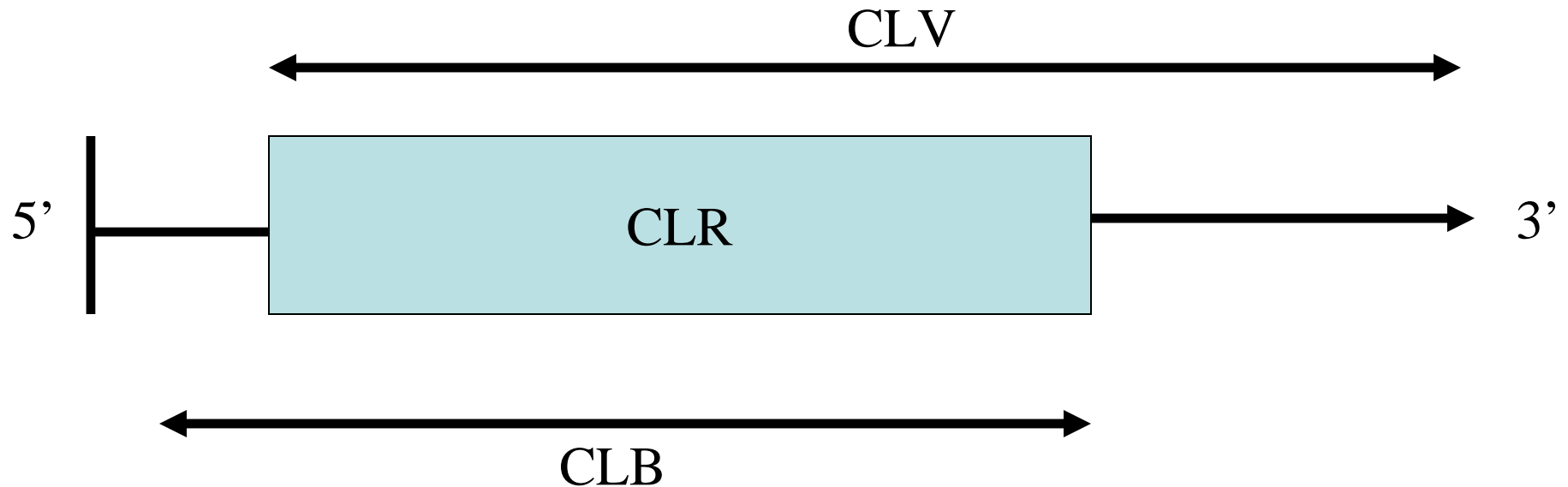
* validated against finished Chrom. 21 sequence

- The clones cover the genome 38.7X times
- Data is from 5 individuals (roughly 3X, 4 x .5X)

Chromatogram Base Calling



A sequence of basecalls is generated by mapping the recorded peaks to an idealized trace by omitting some peaks, and splitting others.



Trimming identifies the regions of good quality for the assembler to use (CLR), as the intersection of the region free of vector (CLV) and the region free of bad quality (CLB).



runCA Pipeline



1. Create Stores
 - gatekeeper
 - PopulateFrgStore
2. Find Repeats
 - meryl
3. Overlap
 - overlap
 - grow-overlap-store
4. Error Correction
 - correct- frags
 - correct-olaps
 - update-erates
5. Unitigging
 - unitigger
 - consensus -U
6. Scaffolding
 - cgw
 - consensus
7. Finalize Data
 - Terminator
 - qc file



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Assembly Stores



- `asm.gkpStore` - name-id mapping, mate pairs
 - populated by gatekeeper
 - dump with `dumpGatekeeper` (output in `STDERR`)
- `asm.frgStore` - bases, qualities, clear range
 - populated by `PopulateFragStore`
 - dump with `dumpFragStore`
- `asm.ovlStore` - overlaps between reads
 - populated by `grow-olap-store`
 - dump with `dump-olap-store`



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Meryl: k-mer statistics



Frequent k-mer statistics: asm.mers

```
count >325
22-mer sequence AAAGCCCAAAGCCCAAAGCCCA
>228
AACAGCTCGATCACGTCGCTGT
```

How much of the DNA is in 300 copies or more?

```
% grep '>' asml.mers | sed 's/>/' | awk '{if ($1>300) sum+= $1} END {print sum;}'
```

**Not every repeat is mis-assembled,
but repeats cause (almost) every mis-assembly.**



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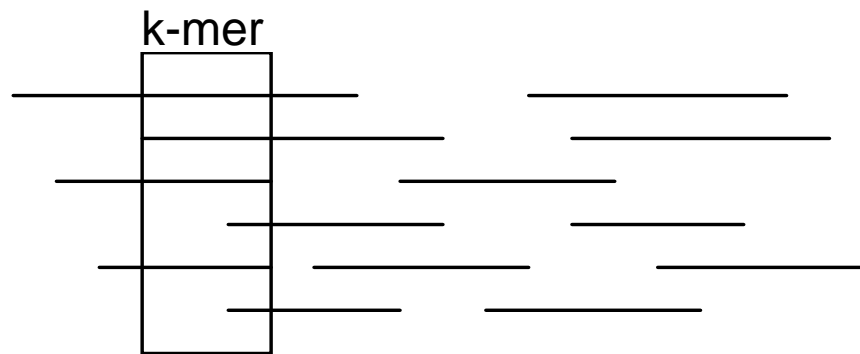
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All pairs alignment

- Needed by the assembler
- Try all pairs – must consider $\sim n^2$ pairs
- Smarter solution: only $n \times$ coverage (e.g. 8) pairs are possible
 - Build a table of k-mers contained in sequences (single pass through the genome)
 - Generate the pairs from k-mer table (single pass through k-mer table)



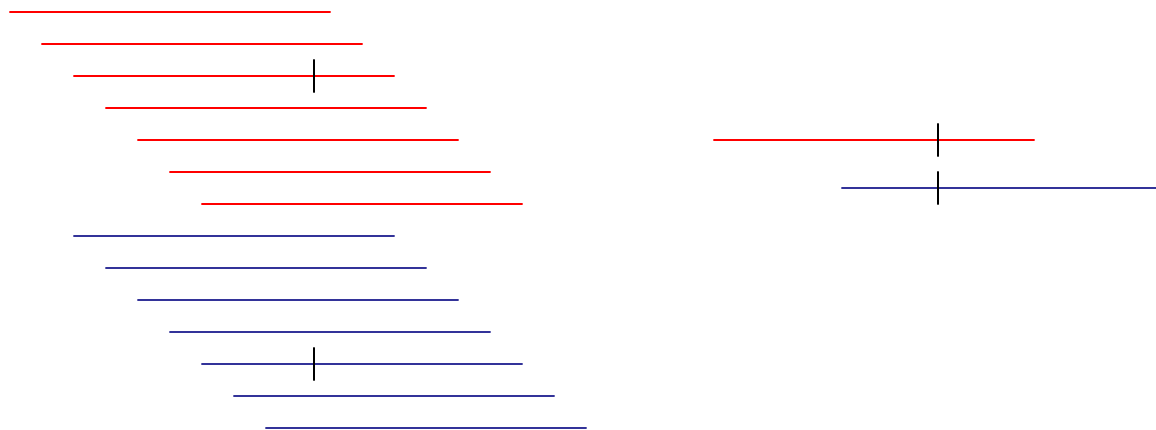
Overlapper

- Find all overlaps $\geq 40\text{bp}$ allowing 6% mismatch.
- Use k-mer ($k=22$) seed matches with $O(nd)$ extension where extension quits when probability of seeing given # of errors for amount of sequence aligned is less than 1 in a million.
- Avoid seeding overlaps with k-mers whose occurrence ≥ 100 in the trimmed read set.
- Multiple threads & multiple instances allowed depending on the input size.



Overlapper & Screening

- High copy repeats are filtered by excluding high copy (≥ 100) 22-mers as seeds.
- **Warning:**
Sequencing error can accidentally cause low copy number seeds in high copy repeat regions creating low coverage unitigs of collapsed repeats.





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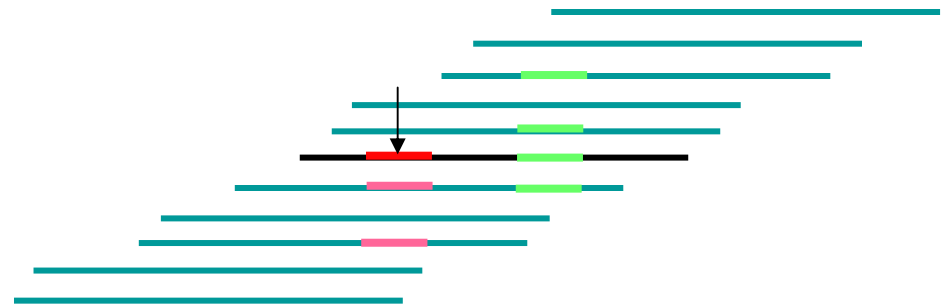
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If a k-mer ($k=10$) matches a k-mer from an overlapping read then the bases in the k-mer of the read are confirmed.

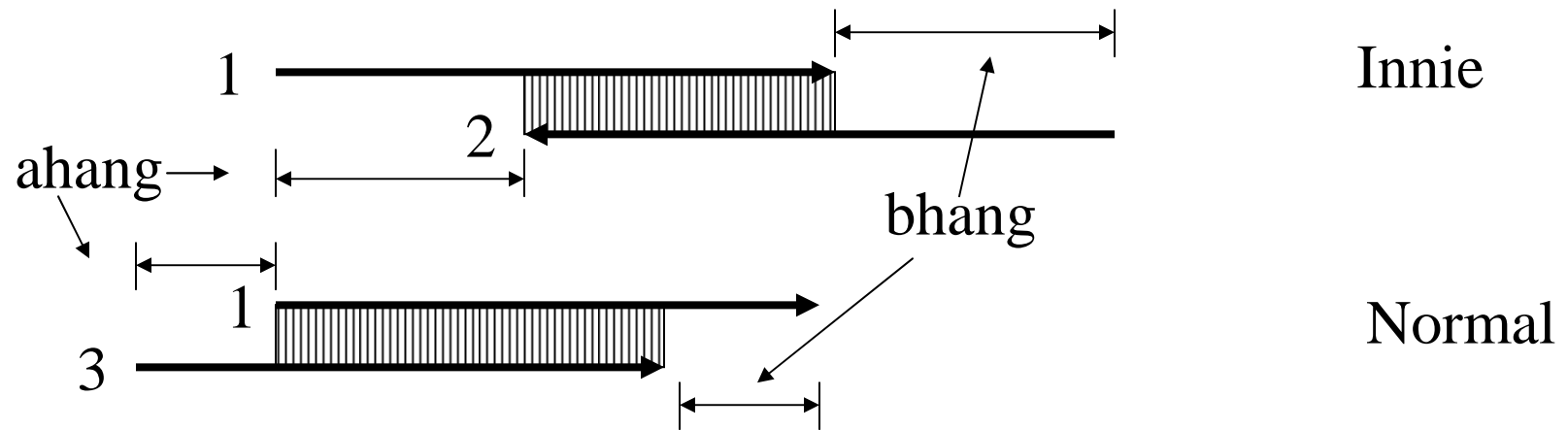
If a **base** is not confirmed and the 1-neighborhood of an overlapping k-mer matches it then there is a vote for correction. The majority correction vote is applied to the sequence.

Note: Sequences are not actually changed, only overlaps are re-evaluated as single base pair errors are “corrected”.



```
ACGTACCGATATGACAC
ACGTACCGTTATGACAC
ACGTACCGATATGACAC
ACGTACCGATATGACAC
```

dump-olap-store



1	2	I	ahang	bhang	1.3	0.1
1	3	N	-ahang	-bhang	2.3	0.4

Original Error Rate

After Error Correction



Overlap degrees



8x coverage: each read overlaps approx. 8 reads off of each end

$a_{hang} < 0$ - overlap off of 5' end

$b_{hang} > 0$ - overlap off of 3' end

```
% awk '{if ($4 < 0) end5++; if ($5 > 0) end3++;} END {print end5, end3}}' asm.overlaps
```

$end5 \text{ overlaps} > end3 \text{ overlaps}$ - normal (3' end is “dirtier”)

$end5 \text{ overlaps} < end3 \text{ overlaps}$ - possible vector trimming problem

```
% awk '{print $1}' asm.overlaps | sort -u | wc -l - # reads with overlaps
```

many reads w/o overlaps - trimming problem or ubiquitous repeat



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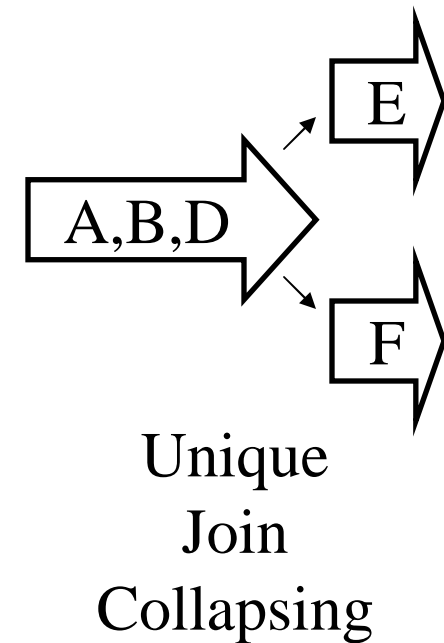
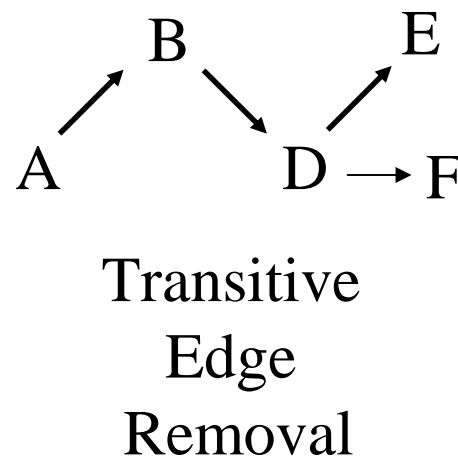
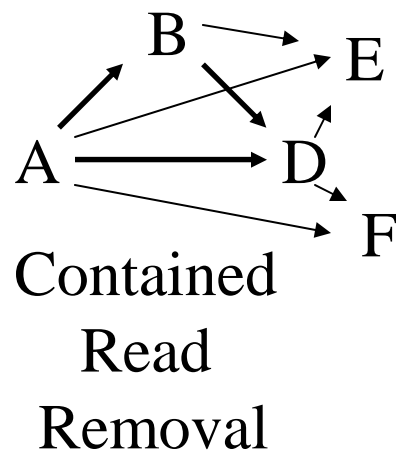
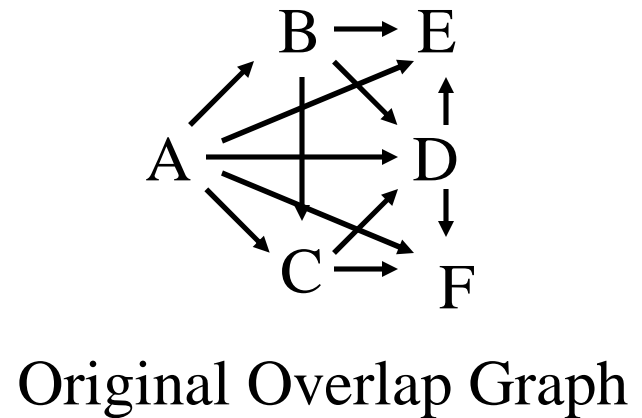
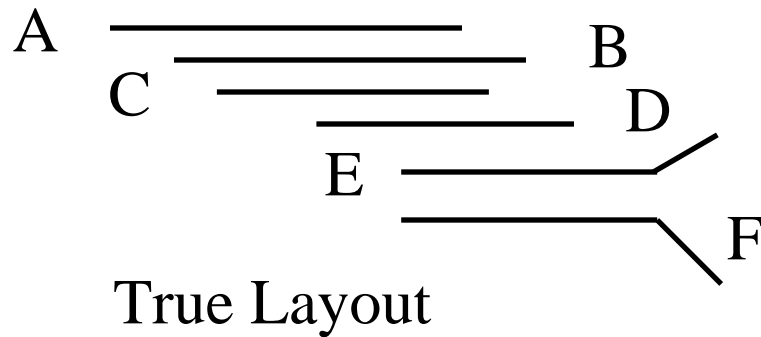
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Unitigging



Theorem: SCS of unitigs = SCS of reads

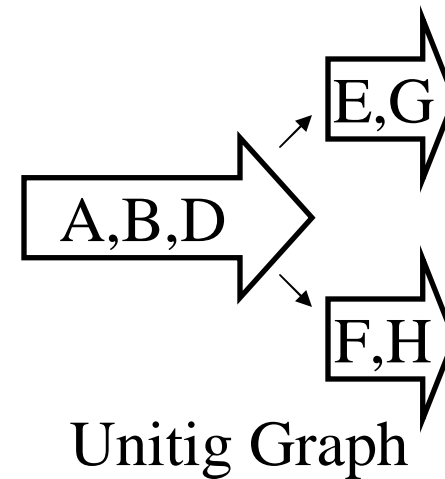
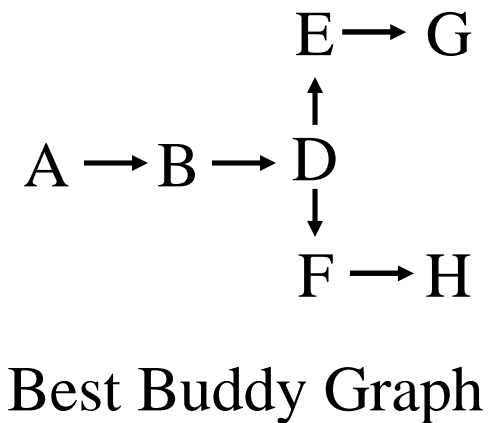
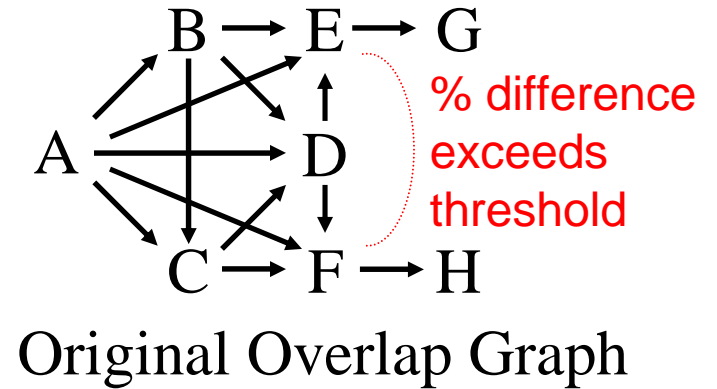
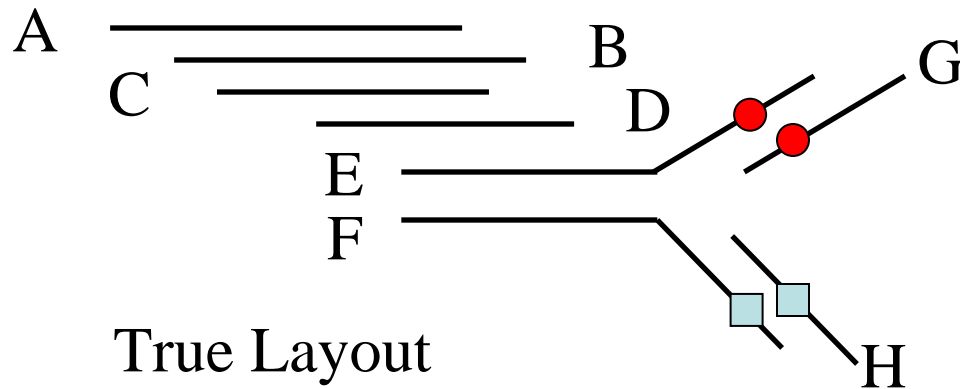


Revised Unitigging



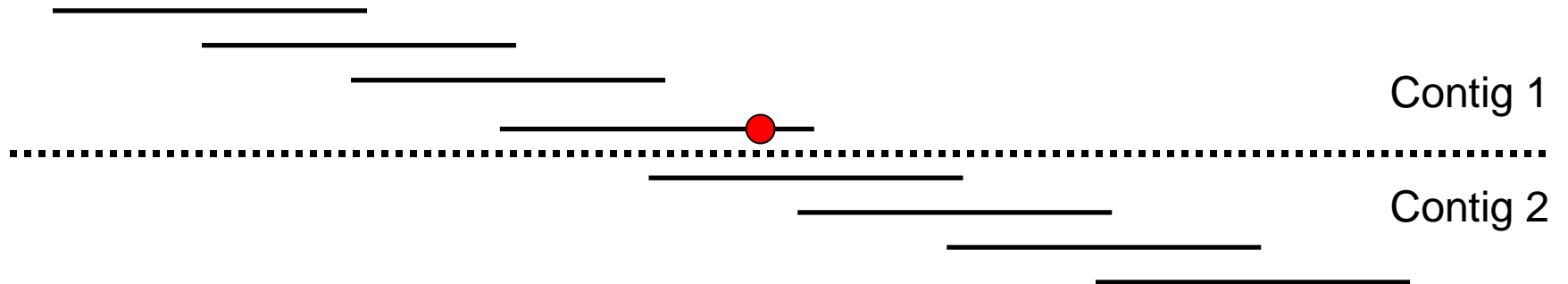
- Exact Unitigging is computationally expensive
- Instead CA unitigger finds the “best” overlap on each end of each read—its “best buddy”.
- Unitigs are chains of mutually unique best buddies—adjacent reads are best buddies of each other and of no other read.
- This takes time and space linear in the number of reads.
- In rare cases results are different from graph reduction.
 - Low coverage regions
 - High fidelity repeat copies

Best Buddy Unitigging



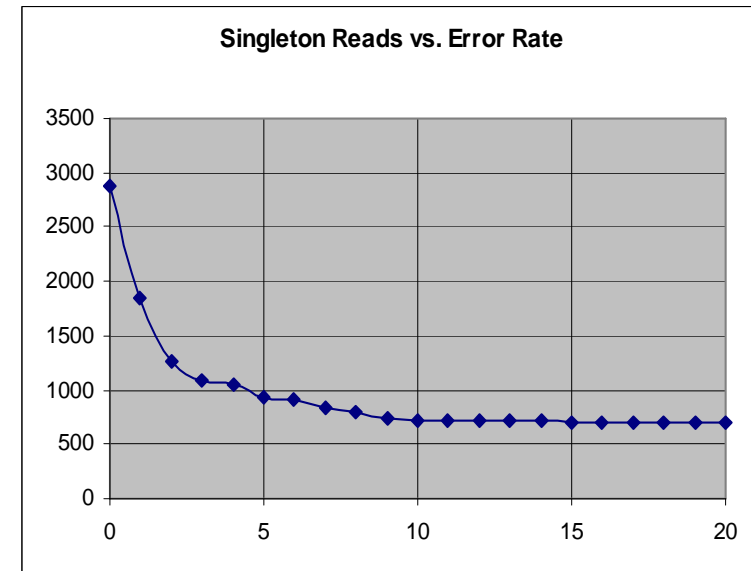
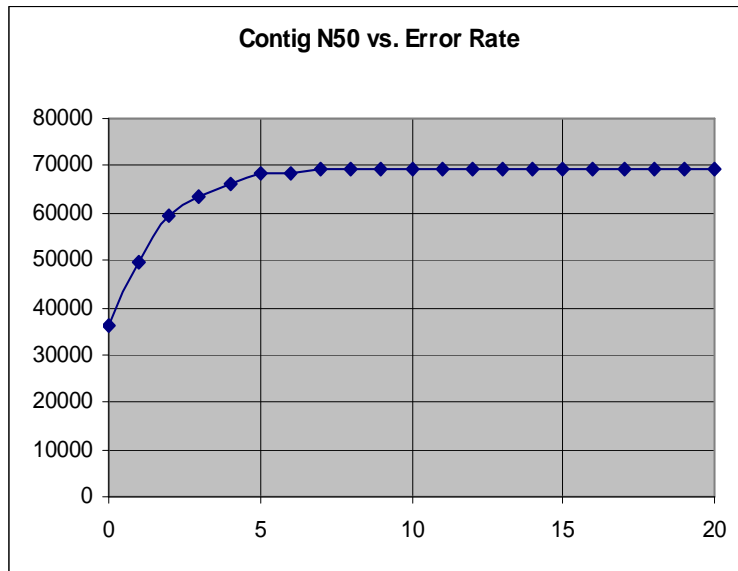
Threshold set with unitigger -e (ERATE, utgErrorRate)

False Negatives: Sequencing Error, Trimming



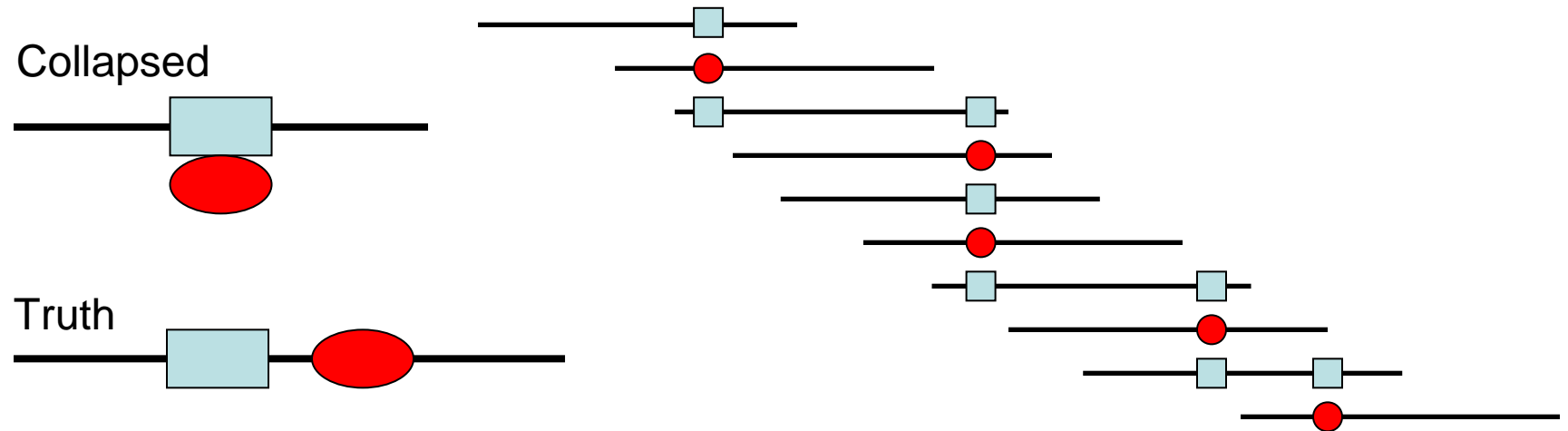
- Overlaps are “missed” if the overlapping basecalls have sequencing error beyond the threshold.
- Assembly is fragmented into smaller chunks, or reads left as singletons.

Sequencing Error Effect



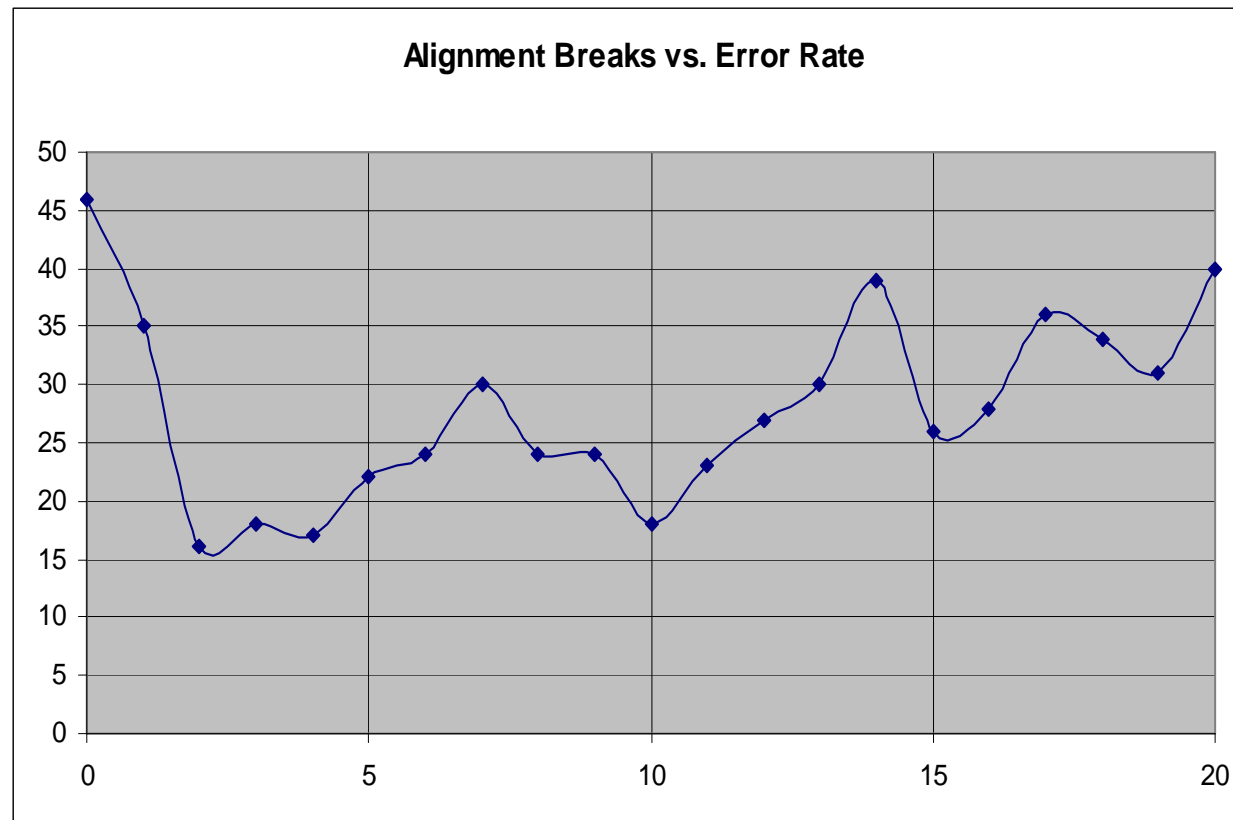
In general, contigs get larger and more reads are placed as the error rate threshold is increased.

False Positives: Repeats



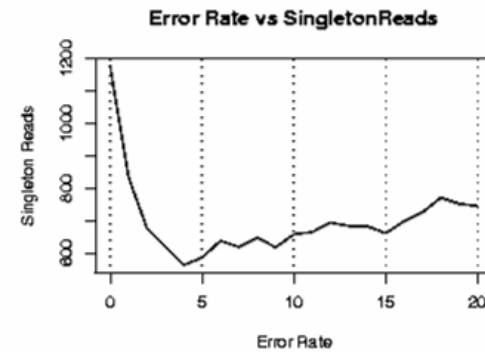
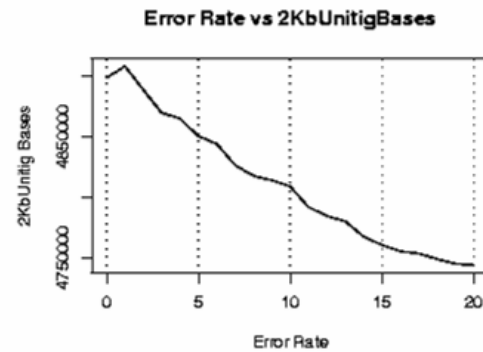
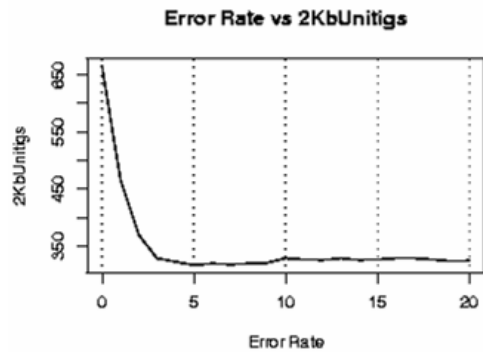
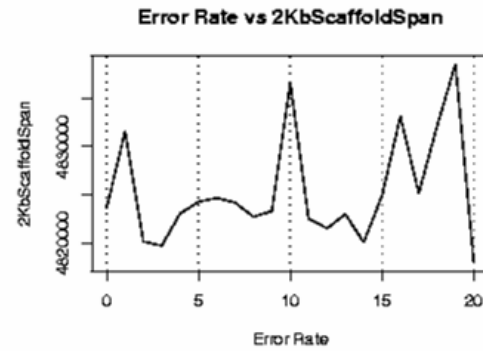
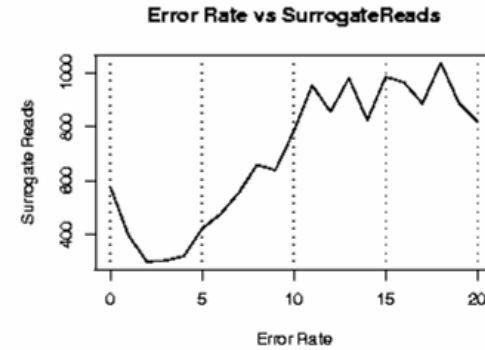
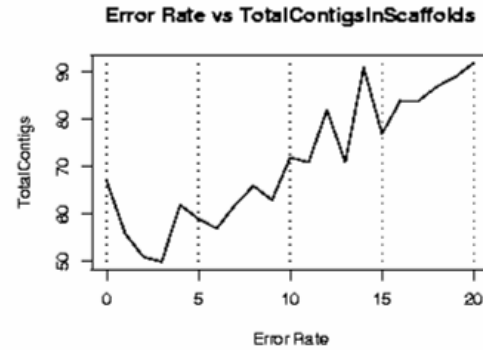
- Reads originating in different copies will “falsely” overlap if % difference between repeats is less than threshold.
- Genome is mis-assembled as reads from different repeat copies are collapsed together as the unitigger becomes less sensitive to slight differences between repeats.

Repeat Effect

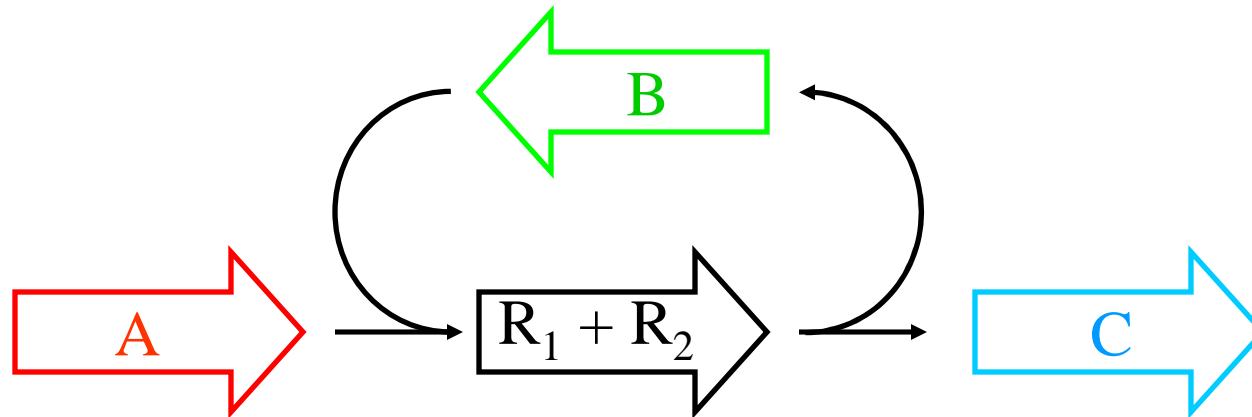
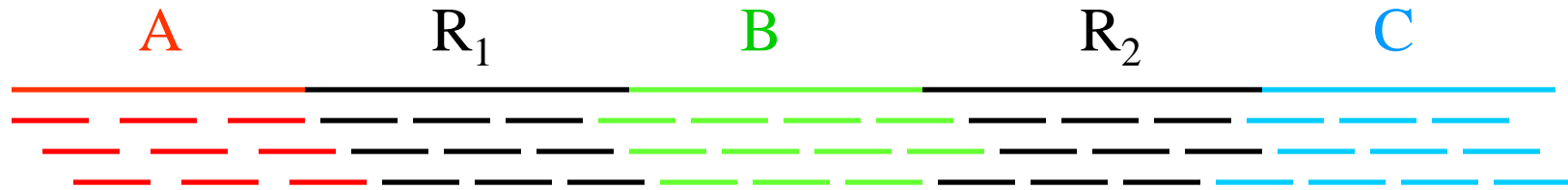


In general, more repeats are mis-assembled as the error rate threshold is increased.

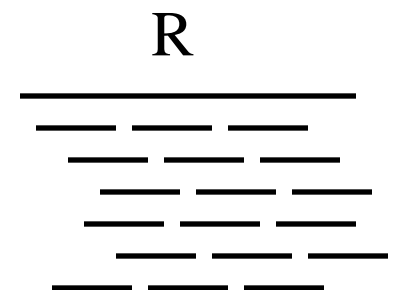
Unitig Error Rate Impact



Unitig Scoring

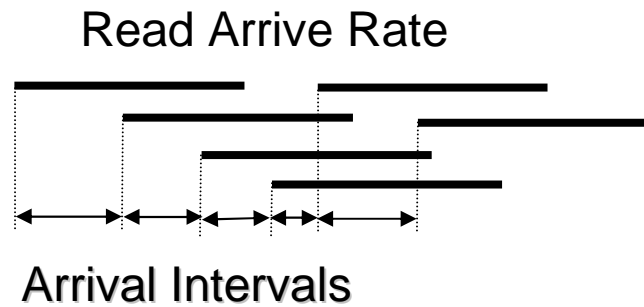


The arrival rate of reads within repeat unitig R is statistically higher than for unique unitigs A, B or C. The corresponding A-stat will mark the unitig as unreliable.



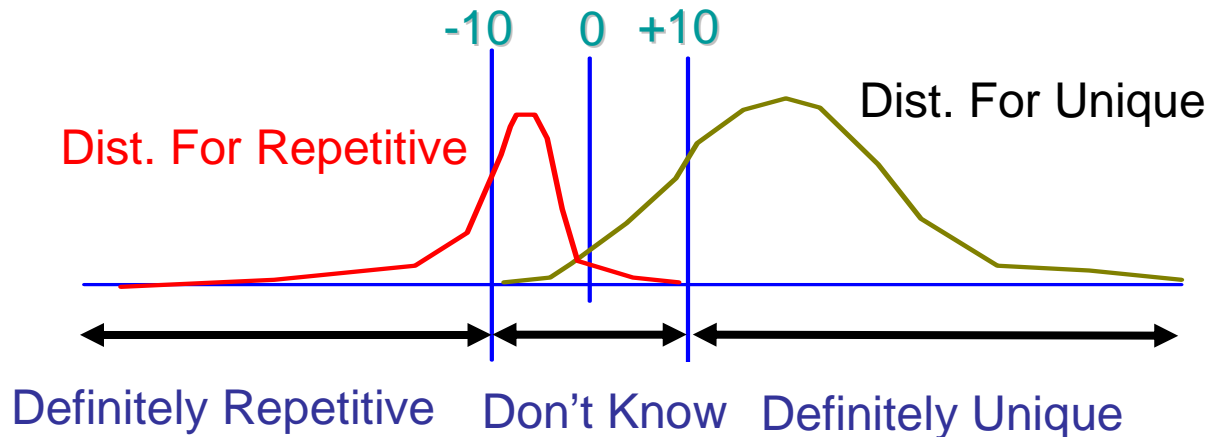
Note: Requires uniform distribution of reads.

Identifying Unique DNA



Expected Coverage is:
(Sum of read lengths) /
Genome Size

Discriminator A-Statistic is log odds ratio of probability unitig is for unique DNA versus 2-copy DNA.



Correct for biases:

- `cgw -j (ASTAT)` : set threshold for definitely unique
- `unitigger -l (utgGenomeLen)` : adjust genome size estimate, boost borderline unitigs



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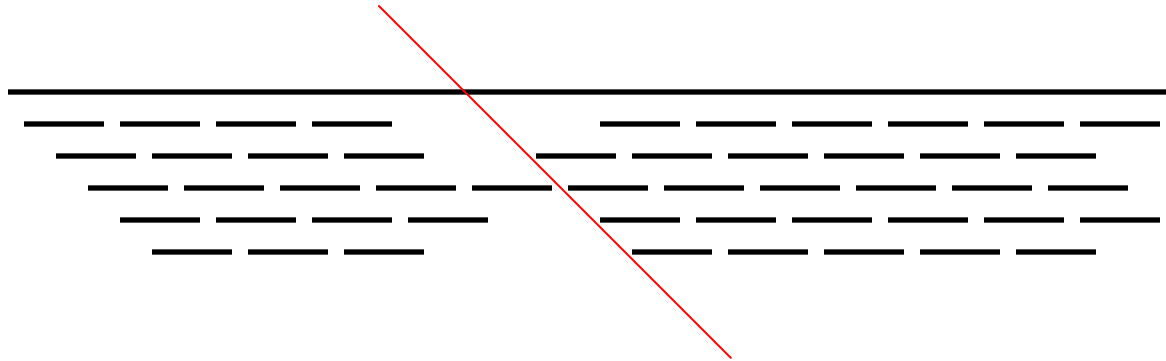
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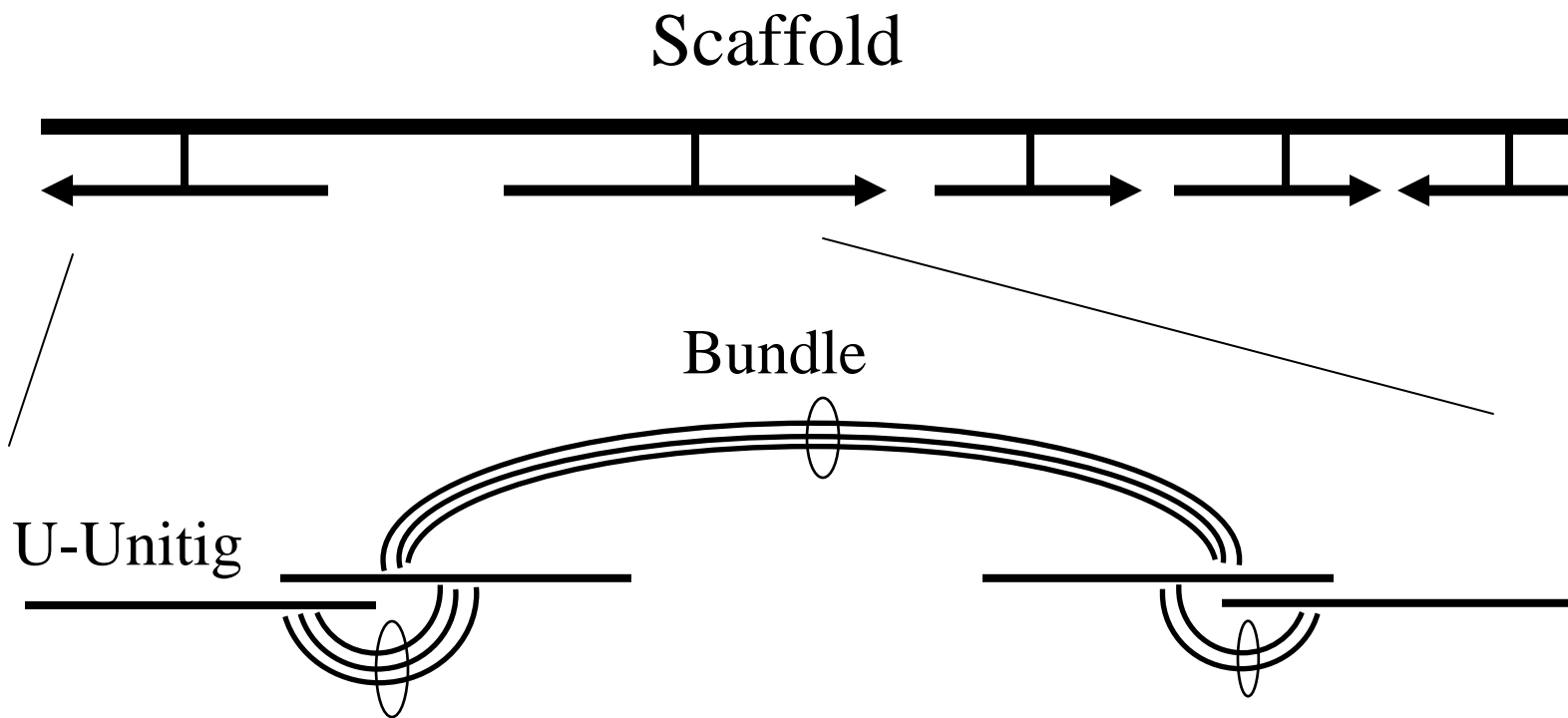
Unitig Splitting



Unitigs are split when the coverage level drops below a threshold, and there are no mates connecting the unitig.

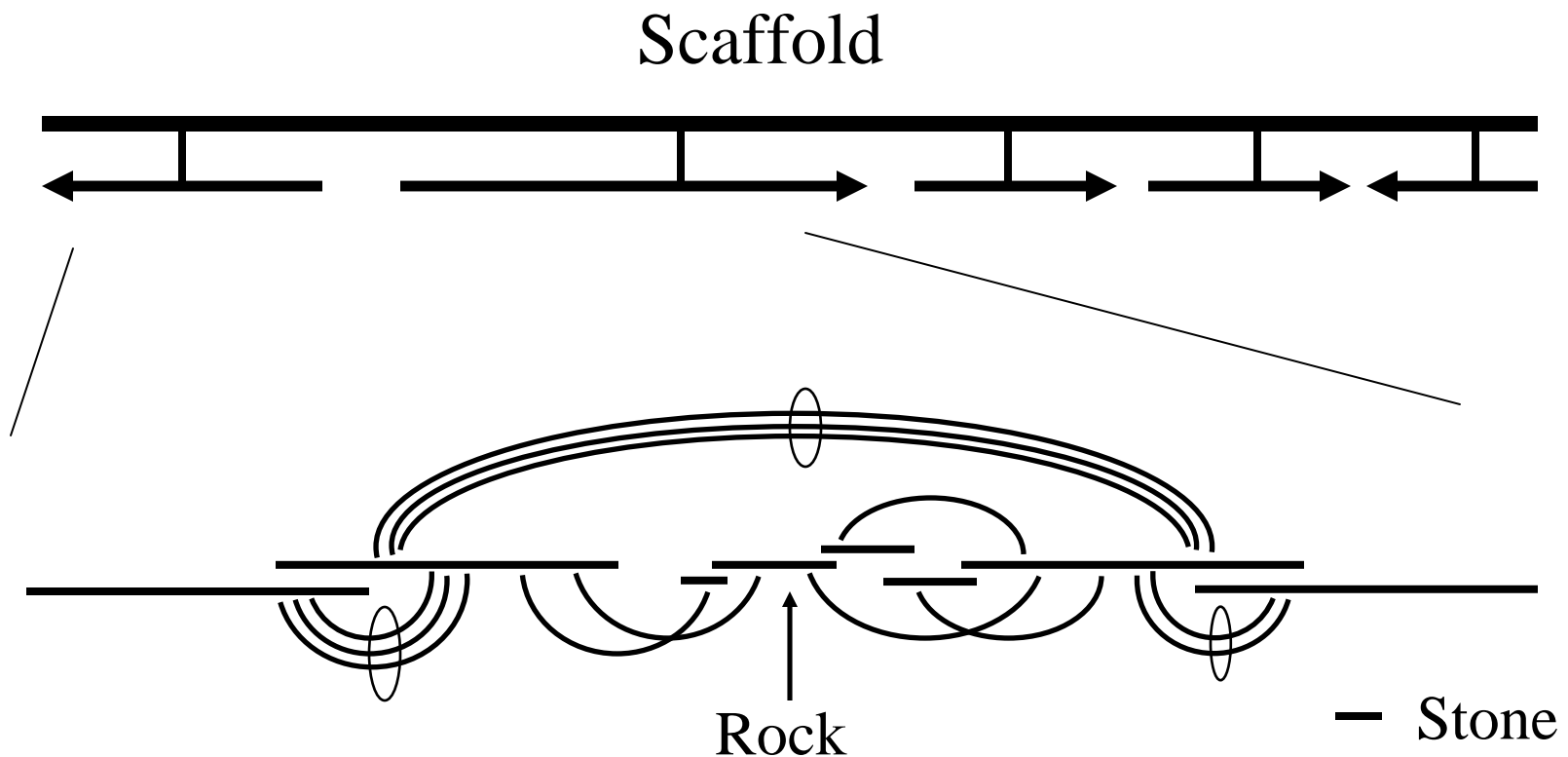
After this step, unitigs are opaque, and every read will be placed in exactly one unitig.

Initial Scaffolding



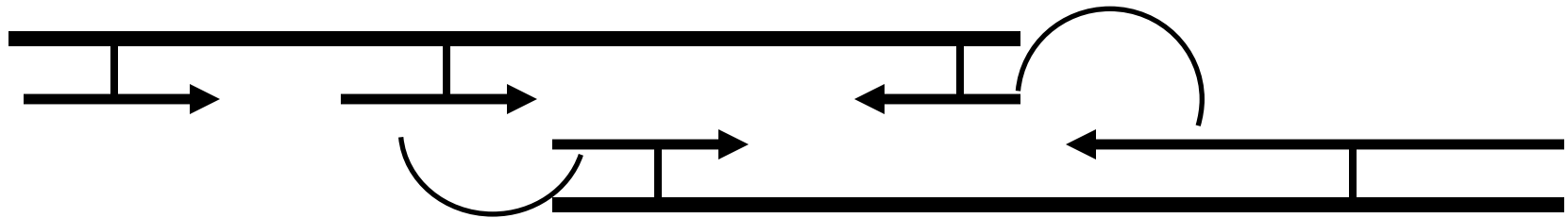
Create a initial scaffold of unique unitigs (U-Unitigs) whose $A\text{-stat} > 5$. Also recruit borderline unitigs whose $A\text{-stat}$ is > 2 and have consistent mates with the U-Unitigs.

Repeat Resolution



Place rocks (A-stat > 0 with multiple consistent mates), and stones (single mate and overlap path with placed objects) into the gaps. Pebbles, unitigs lackings mates, are no longer incorporated regardless of overlap qualities.

Scaffold merging



After placing borderline unitigs and rocks, there may be sufficient mates to merge scaffolds (mates from stones are not considered). If multiple orientations are possible, choose the scaffold merge with the happiest mates.

This in turn may allow for new rocks and stones to be placed, so iterate these steps until the scaffold stabilizes.



Mate Bundling



- The CA scaffolder requires accurate library size estimates.
- Generally necessary to run scaffolder at least twice.
- CGW outputs revised library sizes, repeat until convergence.
- May need to manually split libraries if distributions are multi-modal.

Assembly Dregs

- **Degenerate unitigs** are unitigs with poor A-stat values and not in any scaffold as a rock or stone. (Single contig/unitig scaffolds with a good A-stat are acceptable).
- **Non-unique surrogate unitigs** are unitigs incorporated as stones in multiple places in the scaffold. Consequently, their reads will be multiply placed.
- **Scaffolding Merging** is not done with stones or degenerates so scaffolds may end even though there are unambiguous mates links to follow.



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Assembler outputs



asmb1.asm - all the information in Celera message format

asmb1.qc - summary statistics

asmb1.fasta, asmb1.contig - all the contigs, surrogates and degenerates

asmb1.placed.fasta, .contig - all the contigs

asmb1.surrogates.fasta, .contig - all the surrogates

asmb1.degenerates.fasta, .contig - all the degenerates

asmb1.singletons - all the singletons

asmb1.scaffolds.fasta - all the scaffolds, 60 Ns replace the gaps

asmb1.scaffolds.info - contig order/orientation for scaffolds



The .qc file



```
[Scaffolds]
TotalScaffolds=2
MeanContigsPerScaffold=23.50
MaxContigsPerScaffold=30

TotalBasesInScaffolds=3298141
MeanBasesInScaffolds=1649070.50
MaxBasesInScaffolds=2100614
N50ScaffoldBases=2100614

TotalSpanOfScaffolds=3310522
MeanSpanOfScaffolds=1655261.00
MaxScaffoldSpan=2104833
IntraScaffoldGaps=45
MeanSequenceGapSize=275.13

[Top_5_Scaffolds_contigs_size_span_avgContig_avgGap]
0=30 2100614 2104833 70020.47 145.48
```

<http://www.cbcbl.umd.edu/research/castats.shtml>

50% of genome is in contigs larger than N50

Example:

1 Mbp genome

Contigs: 300, 100, 50, 45, 30, 20, 15, 15, 10,

N50 size = 30 kbp

$$(300+100+50+45+30 = 525 \geq 500\text{kbp})$$

Note:

N50 is meaningful for comparison only when genome size is the same



Assembly Quality



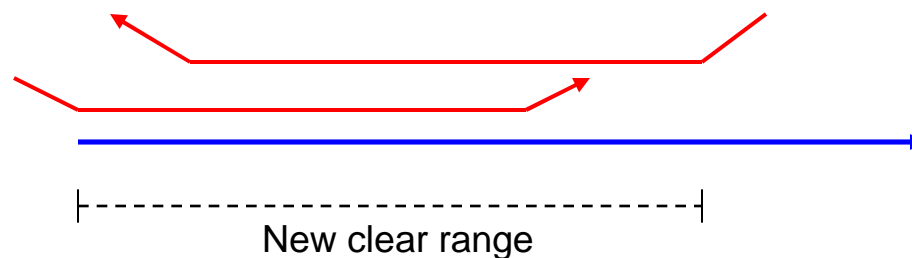
- AMOS Validation Tools
 - Library Construction
 - Contaminate Sequences
 - Read Trimming
 - Coverage Levels
 - A-stat problems / Degenerate Contigs
 - Local Mis-assembly
- Be aware of potential size/quality tradeoffs.



runCA-OBT Overlap-Based-Trimming



- Find local alignments (“partial overlaps”) between untrimmed reads.
- Use overlapping alignment regions to set new clear range.
- Patterns of overlap forks can automatically find and trim unknown vector sequences.
- runCA-OBT is a work in progress at Venter Institute
 - Does several advanced operations as well: extendClearRanges, resolveSurrogates, resizes Libraries
 - `wgs-assembler/src/AS_RUN/runCA-OBT/doc.tex`





Celera Assembler Summary



- **Strategy**
 1. Compute Overlaps between reads
 2. Simplify Overlap Graph into Unitigs
 3. Score Unitigs based on Coverage
 4. Create Contigs & Scaffold of Unique Unitigs
 5. Fill in gaps with repetitive unitigs
- **Complications**
 1. Vector & Quality trimming to find all overlaps
 2. Unitig Error Rate to separate repeat copies
 3. Unitig Scoring (A-stat) to build contigs from unique pieces



Current Development



- **UMd / CBCB**
 - Overlapping, Repeat Resolution

Steven Salzberg
Art Delcher
- **UMd / IPST**
 - Error Correction, Unitigging

Jim Yorke
- **Venter Institute**
 - OBT, Scaffolding, Consensus

Granger Sutton
- **TIGR**
 - Code Engineering, Bug Fixes

Martin Shumway
Jason Miller